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TWO NEW IRIDOIDS FROM *PATRINIA SCABRA*

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From the roots of *Patrinia scabra* two new iridoids, 3-methylbutyric acid 7-hydroxy-7-hydroxymethyl-4-(3-methylbutyryloxymethyl)-6-oxo-1,6,7,7a-tetrahydrocyclopenta[c]pyran-1-yl ester (**1**) and 6-hydroxy-7-methylhexahydrocyclopenta[c]pyran-3-one (**2**) have been isolated. Their structures were determined by means of NMR spectra and X-ray crystallographic analysis.

Keywords: *Patrinia scabra*; Iridoids

INTRODUCTION

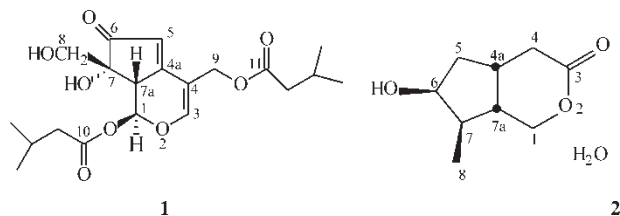
Patrinia scabra Bunge is a wild plant growing mainly in the northeastern part of China. The plant is used as a traditional medicine to treat leukemia, cancer and for regulating host immune response in China. Some iridoids and iridoid glycosides have been found previously in this plant [1,2]. We report here the isolation and structural elucidation of two new iridoids, 3-methylbutyric acid 7-hydroxy-7-hydroxymethyl-4-(3-methylbutyryloxymethyl)-6-oxo-1,6,7,7a-tetrahydrocyclopenta[c]pyran-1-yl ester (**1**) and 6-hydroxy-7-methylhexahydrocyclopenta[c]pyran-3-one (**2**), from the roots of *Patrinia scabra*.

RESULTS AND DISCUSSION

By ethanolic extraction of the air-dried roots of *P. scabra*, an EtOAc-soluble fraction was obtained and further isolated by silica gel chromatography to afford two iridoids (Fig. 1).

Compound **1** was obtained as white needles, $[\alpha]_D^{25} -196.3$ (*c* 1.0 in MeOH), mp 92–93°C. UV $\lambda_{\text{max}}^{\text{MeOH}}$ nm (log ϵ): 302 (4.14). The molecular formula, C₂₀H₂₈O₈, was established by the $[M + Na]^+$ peak at *m/z* 419 in the FAB-MS spectrum; EI-MS *m/z*: 396[M⁺]. The IR spectrum indicated the presence of an hydroxy group (3276 cm⁻¹), two ester carbonyl groups (1750 and 1735 cm⁻¹) and a conjugated ketone carbonyl group (1683 cm⁻¹). The ¹H and ¹³C NMR spectra of **1** (Table I) showed four secondary methyl proton signals at δ 0.92–1.01, two methine carbon signals at δ 25.6 and 25.7, two methylene

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FIGURE 1 Structures of compounds **1** and **2**.

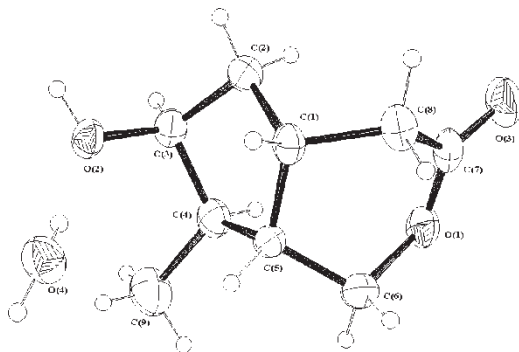
carbon signals at δ 42.1 and 43.0 as well as two carbonyl carbon signals at δ 170.4 and 172.9, indicating the presence of two isovaleryloxy ester moieties, which were further confirmed by 2D COSY of **1**. The remaining 10 carbon signals gave rise to two oxy-methylenes (δ 65.1 and 59.8), two sp^3 -hybridised methines (δ 92.4 and 42.1), two sp^2 -hybridised methines (δ 154.2 and 118.5) as well as four quaternary carbons (δ 75.1, 102.3, 164.8 and 207.3), including a conjugated ketone, which yield a cyclopenta[*c*]pyran-type iridoid structure for **1**. The 1H NMR spectrum of **1** also showed a singlet at δ 7.03 (H-3) and a doublet at δ 5.95 (H-5) from the two trisubstituted double bonds located at C-3-C-4 and C-4a-C-5. Consequently, the conjugated ketone carbonyl group was located at C-6. The presence of a CH_2OH group was confirmed by a signal at δ 3.82, 65.1. The signals at δ 6.39 (1H, d, $J = 11.0$ Hz, H-1) and 3.08 (1H, dd, $J = 2.3, 11.0$ Hz) indicated the α configuration of H-1 and β configuration of H-7a [3].

The relative stereochemistry for **1** was established by NOESY experiments which showed a correlation peak between H-7a and the CH_2OH group, indicating that the CH_2OH group is attached to C-7 with the β configuration. The absolute stereostructure has not been determined yet.

Compound **2** was also obtained as white needles, mp 98.5–100°C; EI-MS m/z : 170 [M^+], 152, 139, 126, 97, 81, 69 (base), 55. The IR spectrum showed a hydroxyl absorption at 3348 cm^{-1} and a lactone carbonyl at 1733 cm^{-1} . The ^{13}C NMR spectrum of **2** (Table I) exhibited 9 carbon signals which were resolved into one methyl, three methylenes, four methines and a lactone carbonyl with the help of DEPT experiments. The 1H – 1H COSY spectrum showed the connectivities of the proton coupling sequence for the C-4-C-4a-C-5-C-6-C-7-C-7a-C-1 fragment. It also showed correlation peaks of H-4a with H-7a, and H-8

TABLE I NMR data of compounds **1** and **2** in $CDCl_3$

Position	1		2	
	δ_C	δ_H (J_{Hz})	δ_C	δ_H (J_{Hz})
1	92.4	6.39 (d, 11.0)	68.58	4.15 (dd, 11.7, 3.5) 4.33 (dd, 11.7, 4.0)
3	154.2	7.03 (s)	173.70	
4	102.3		34.41	2.37 (dd, 15.0, 4.0) 2.65 (dd, 15.0, 7.2)
4a	164.8		32.52	2.94 (m)
5	118.5	5.95 (d, 2.3)	41.48	1.40 (m), 2.04 (m)
6	207.3		75.28	4.11 (m)
7	75.1		41.55	1.91 (m)
7a	42.1	3.08 (dd, 2.3, 11.0)	41.38	2.17 (m)
8	65.1	3.82 (s)	12.57	1.08 (d, 6.8)
9	59.8	4.74 (d, 12.6) 4.81 (d, 12.6)		
10	170.4			
11	172.9			

FIGURE 2 X-ray crystal structure for compound **2**.

(methyl group proton signal) with H-7. This yielded a 9C cyclopenta[*c*]pyran-type iridoid structure for **2**. The methine signal at δ 75.28 was clearly due to substitution at C-6 with a hydroxy group. The relative stereochemistry of **2** was elucidated by X-ray crystallographic analysis (Fig. 2); it was discovered that **2** is a monohydrate compound, which was further confirmed by the broad absorption from 3100 to 3467 cm^{-1} in the IR spectrum.

EXPERIMENTAL

General Experimental Procedures

The melting point was determined on a RY-2 (Tianjin Analysis Instrument Factory) and is uncorrected. The IR spectrum was recorded on a Bruker Vector-22. NMR spectra were acquired on a Varian INOVA-400 with TMS as internal standard. EI-MS data were obtained on a Varian Mat-212, by the direct inlet method, and UV data were obtained with a Shimadzu UV-265. Chromatographic column: silica gel H (10–40 μ , Qing Dao Oceanic Chemical Industry), Sephadex LH-20 (Pharmacia). All other solvents were analytically pure.

Plant Material

Roots of *P. scabra* Bunge were collected in 2000 from Henan province (China) and identified by Professor H.C. Zheng, Department of Pharmacognosy, School of Pharmacy, The Second Military Medical University. A voucher specimen has been deposited in the herbarium of this institute.

Extraction and Isolation

The air-dried roots of *P. scabra* (10 kg) were extracted with EtOH (3 \times 20 L) at room temperature. After removal of the solvent under reduced pressure, the extract was suspended in water and then partitioned with light petroleum, EtOAc and *n*-BuOH successively. The EtOAc-soluble part (450 g) was separated and fractionated by silica gel chromatography, with petrol–EtOAc (10:1; 5:1; 2:1), into 25 fractions.

Fraction 9 (10 g) was chromatographed on a silica gel column (5 \times 40 cm), eluted with light petroleum–acetone (10:1) to yield 10 further fractions, the sixth of which was chromatographed repeatedly on silica gel and finally yielded **1** (125 mg) as white needles from light petroleum–acetone (5:1).

Fraction 12 (25 g) was chromatographed on silica gel (8 × 65 cm), using CHCl₃–MeOH (20:1 and 10:1 successfully) to give five fractions, the third of which was subjected to a Sephadex LH-20 column (2 × 100 cm) with CHCl₃–MeOH (1:1) as eluent to afford crude crystals, which were recrystallized from light petroleum–acetone (5:1) to give **2** (38 mg).

Structure and Identification

Compound **1**, white needles, mp 92–93°C, $[\alpha]_D^{25} - 196.3$ (*c* 1.0 in MeOH), HREI-MS *m/z*: 396.1781 (calcd for C₂₀H₂₈O₈ 396.1784). IR (KBr) ν_{\max} (cm⁻¹): 3276 (OH), 1750, 1735, 1683 (C=O). EI-MS *m/z*: 396 [M⁺], 295, 276, 192, 179, 151, 136, 85, 57 (base). ¹H and ¹³C NMR data see Table I.

Compound **2**, white needles, mp 98.5–100°C. HREI-MS *m/z*: 170.0949 (calcd for C₉H₁₄O₃ 170.0943). EI-MS *m/z*: 170 [M⁺], 152, 139, 126, 97, 81, 69 (base), 55. IR (KBr) ν_{\max} (cm⁻¹): 3347 (OH), 2962, 2933, 2903, 2880, 1733 (C=O), 1434, 1389, 1347, 1281, 1187, 1167, 1078, 1054, 1017, 976. ¹H and ¹³C NMR data see Table I.

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